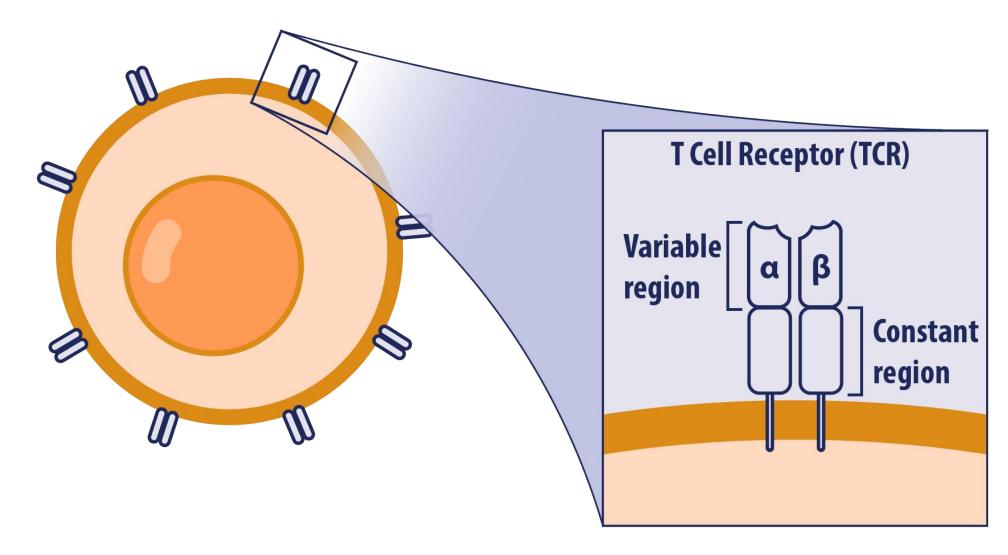
Identifying αβ T cell clones via pooling and b-matching

Tyler Daddio Ion Mandoiu, Ph.D. University of Connecticut

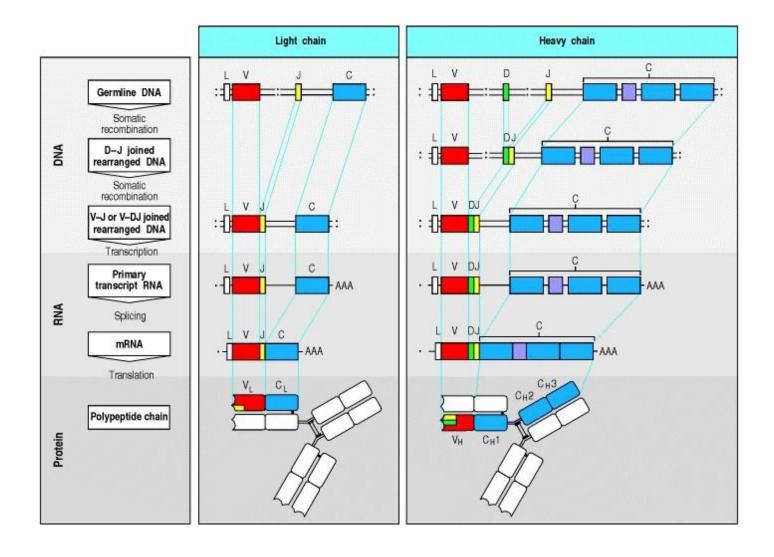
Itinerary

- 1. $\alpha\beta$ T cells and T cell receptors (TCRs)
 - 1. What are they?
 - 2. How are they formed?
- 2. Identifying TCRs
- 3. Reconstructing clones
 - 1. Our approach
 - 2. Results and future work

αβ T cell



How are TCRs formed?



Janeway CA Jr, Travers P, Walport M, et al. Immunobiology: The Immune System in Health and Disease. 5th edition. New York: Garland Science; 2001.

Multiple rearrangements?

- Up to 30% of T cells contain two in-frame α -recombinants (dual α)
 - Only 10% of T cells express both
- Up to 2-10% contain two in-frame β -recombinants (dual β)
 - Only 1-3% of T cells express both

Identifying TCRs: PairSEQ

RESEARCH ARTICLE IMMUNOLOGY

High-throughput pairing of T cell receptor α and β sequences

Bryan Howie^{1,*}, Anna M. Sherwood^{1,*}, Ashley D. Berkebile¹, Jan Berka¹, Ryan O. Emerson¹, David W. Williamson¹, Ilan Kirsch¹, Marissa Vignali¹, Mark J. Rieder¹, Christopher S. Carlson² and Harlan S. Robins^{1,2,†}

Science Translational Medicine 19 Aug 2015: Vol. 7, Issue 301, pp. 301ra131 DOI: 10.1126/scitranslmed.aac5624

¹Adaptive Biotechnologies, Seattle, WA 98102, USA.

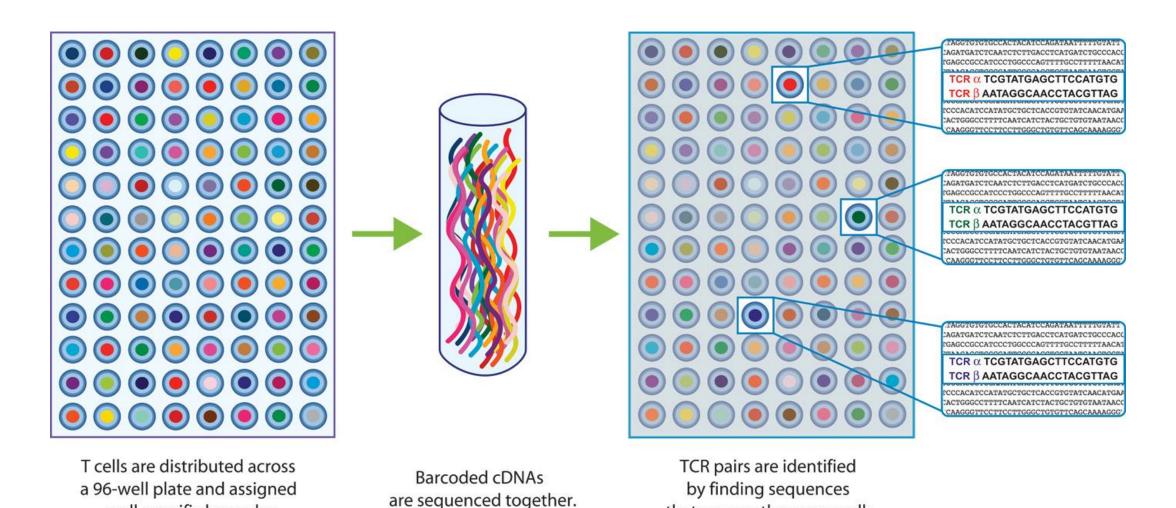
²Fred Hutchinson Cancer Research Center, Seattle, WA 98109, USA.

^{4&}lt;sup>†</sup>Corresponding author. E-mail: hrobins@fredhutch.org

⁺ See all authors and affiliations

Identifying TCRs: PairSEQ

well-specific barcodes.



Howie B, Sherwood AM, Berkebile AD, Berka J, Emerson RO, Williamson DW, et al. High-throughput pairing of T cell receptor α and β sequences. Sci Transl Med. 2015;7(301):301ra131. pmid:26290413

that occupy the same wells.

Benefits

- 1. Can identify hundreds of thousands of TCRs
- 2. Uses existing high-throughput sequencing technologies

Questions

- 1. Is it possible to identify entire clones, not just pairs?
- 2. Can clone recognition be done efficiently?

ALPHABETR



RESEARCH ARTICLE

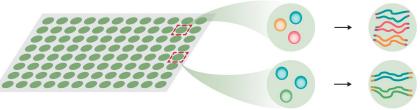
Identifying T Cell Receptors from High-Throughput Sequencing: Dealing with Promiscuity in TCRα and TCRβ Pairing

Edward S. Lee¹, Paul G. Thomas², Jeff E. Mold³, Andrew J. Yates¹*

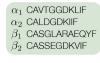
1 Institute of Infection, Immunity & Inflammation, Glasgow Biomedical Research Centre, University of Glasgow, Glasgow, United Kingdom, 2 St. Jude Children's Research Hospital, Memphis, Tennessee, United States of America, 3 Karolinska Institute, CMB, Stockholm, Sweden

ALPHABETR

A T cells are sampled onto 96-well plates at 10-300 cells/well



C Unpaired CDR3α and CDR3β are sequenced in each well



 $lpha_1$ CAVTGGDKLIF $lpha_3$ CAVTYGYLNF $lpha_4$ CALTASGLTF eta_1 CASGLARAEQYF eta_2 CASSHSRYEQYF

CSEVHTARTQYF

- **D** A resampling strategy is used to obtain a list of possible TCR pairs by repeatedly performing steps (i), (ii), (iii)
 - (i) A subset of wells is randomly selected for steps (ii) and (iii)
- (ii) Association scores are calculated for every α and β chain across all wells in the subset

B cDNA libraries are created

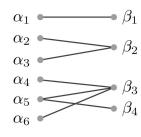
from each well with RT-PCR

$$S_{ij} = \sum_{k=1}^{W} \left(\frac{\delta_{ij}^k}{c_{\alpha}^k} + \frac{\delta_{ij}^k}{c_{\beta}^k} \right)$$

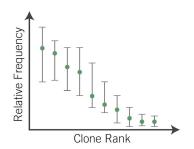
(iii) Scores are used to select likely αβ pairs within each well

	α_1	α_2	α_3	α_4	α_5
β_1	24.0	0.6	1.2	4.3	8.2
β_2	24.0 0.4 1.0 3.2	0.2	60.2	0.7	2.2
β_3	1.0	0.2	1.2	9.0	3.0
β_4	3.2	30.1	0.1	0.4	2.1

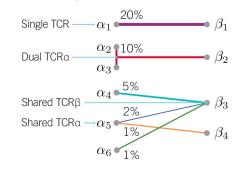
E Pairs from Step D present in more than a threshold proportion of replicates are candidate TCRs



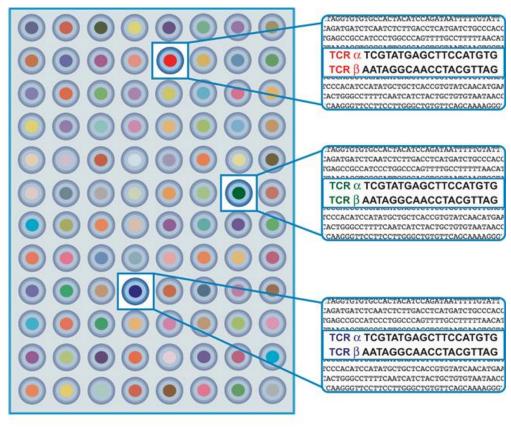
F Clonal frequencies are estimated with maximum-likelihood and used to distinguish β-sharing and dual TCRα



G The output is a list of single and dual TCR clones with their respective clonal frequencies

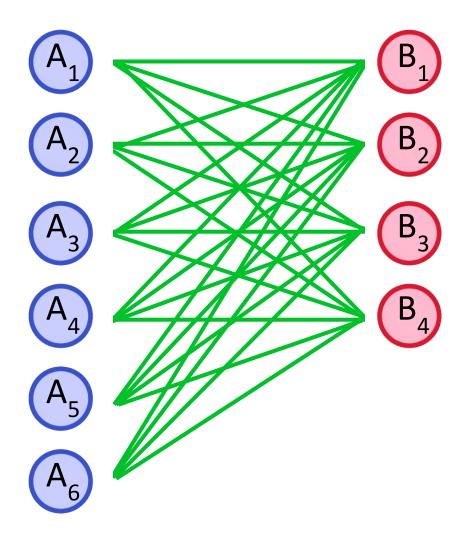


Observations

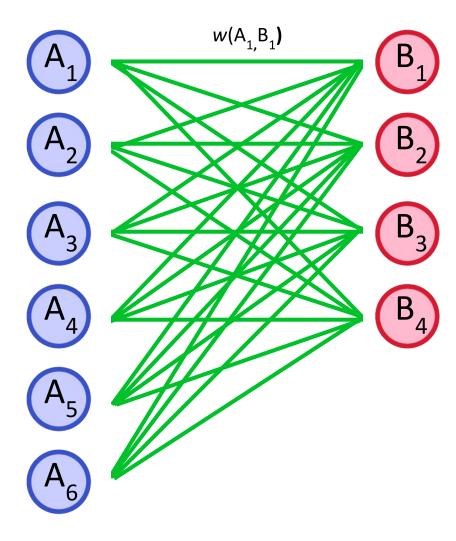


TCR pairs are identified by finding sequences that occupy the same wells.

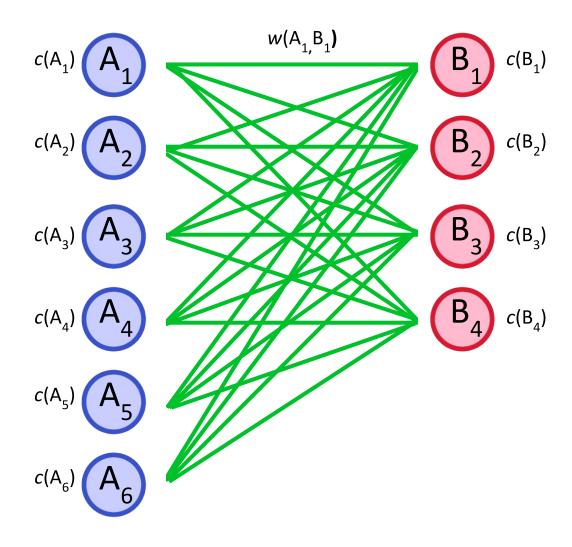
- 1. Many sequences appear in the same subset of wells.
 - These aren't uniquely identifiable!
 - Don't consider sequences directly, match similar well subsets instead
- 2. Modeling dual clones is easy!
 - All recombinants should appear together most of the time
 - Known generative probabilities
 - Up to two recombinants of each type
- 3. Modeling chain-sharing is difficult!
 - Recombinants do not appear together most of the time
 - Unknown generative probabilities
 - Amount of sharing unknown



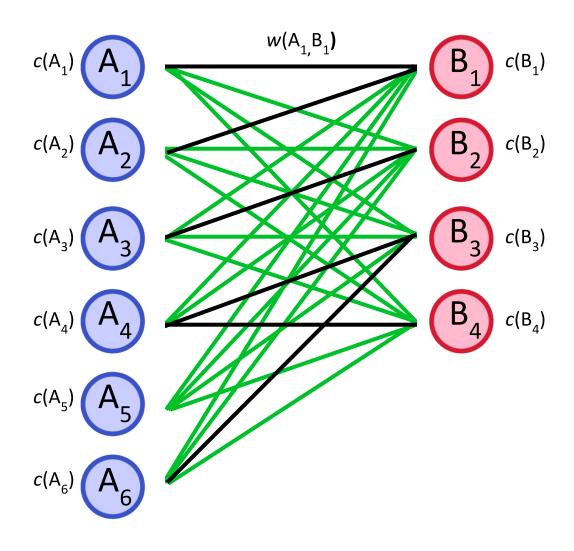
1. Construct a complete bipartite graph where the nodes are the well sets.



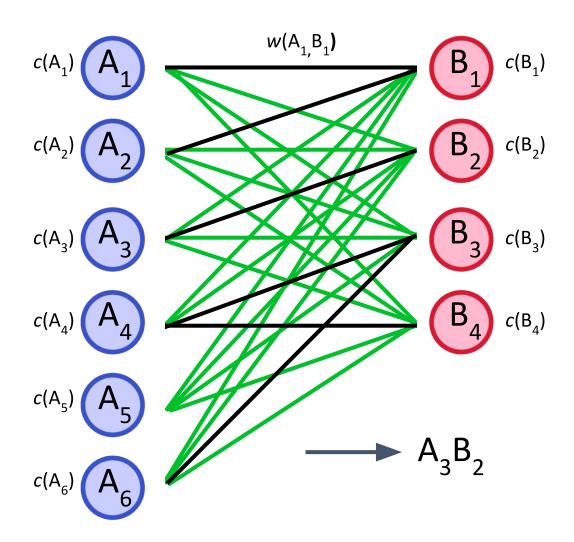
- 1. Construct a complete bipartite graph where the nodes are the well sets.
- 2. Assign a cost w to each edge according to the dissimilarity of the connected nodes.



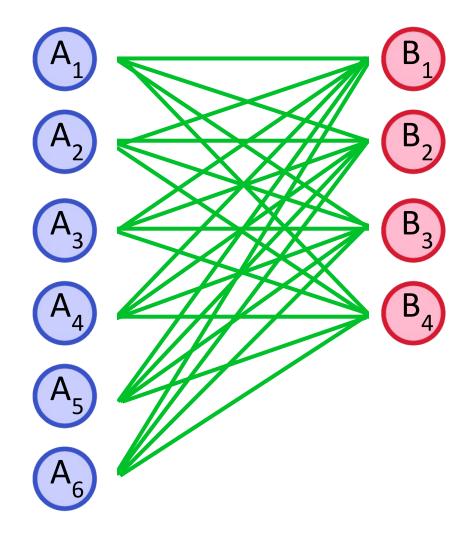
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- 3. Assign a capacity c to each node according to the number of sequences in the corresponding well set.

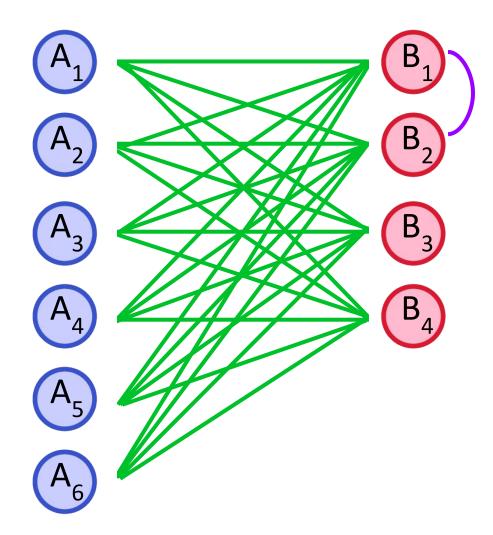


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- 4. Identify a b-matching.

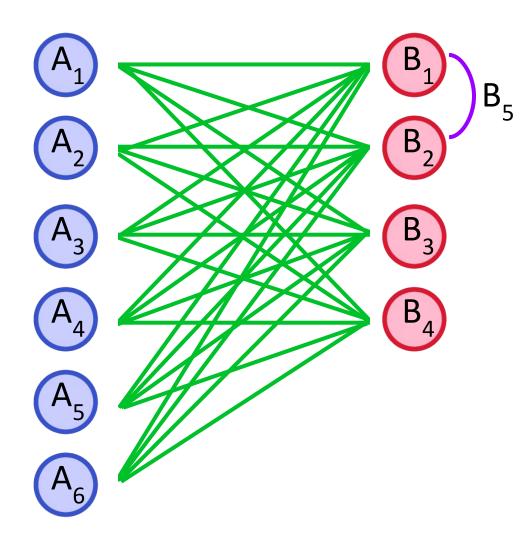


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- 2. Assign a cost w to each edge according to the dissimilarity of the connected nodes.
- 3. Assign a capacity c to each node according to the number of sequences in the corresponding well set.
- 4. Identify a b-matching.
- 5. Output b-matching edges connecting uniquely identifiable well sets.

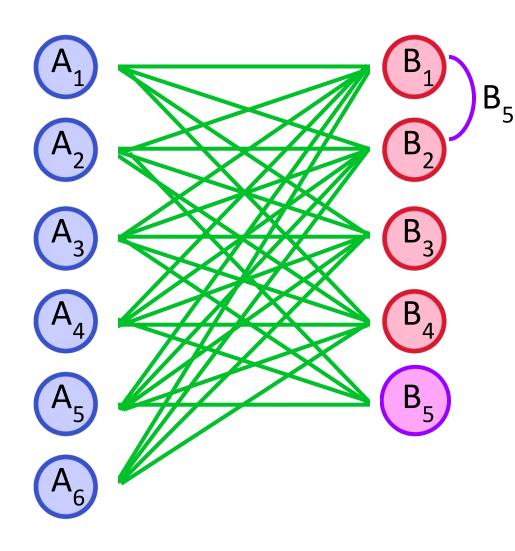




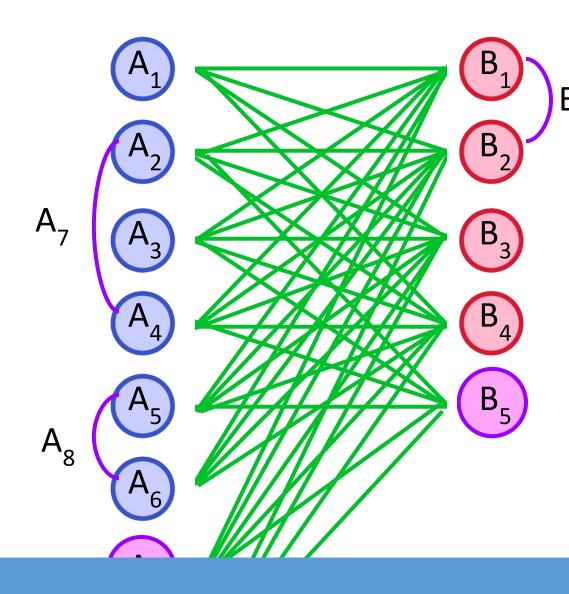
1. Identify pairs of beta well sets that are similar.



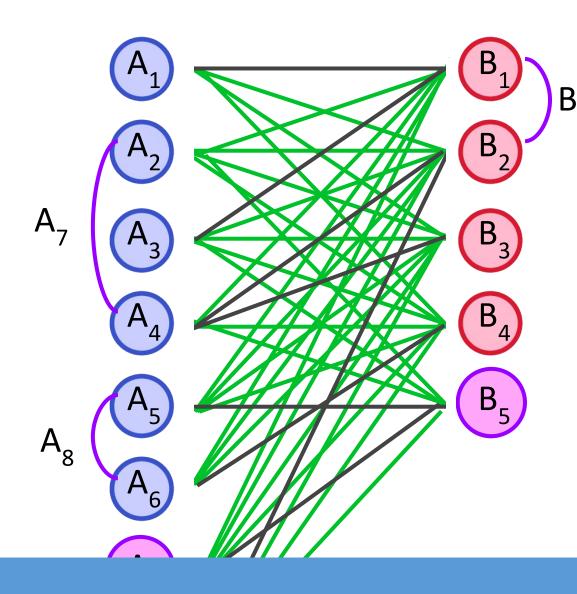
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- 2. Create a new beta well set corresponding to the union of the two individual beta well sets.



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- 1. Identify pairs of beta well sets that are similar.
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- 4. Repeat for alphas.



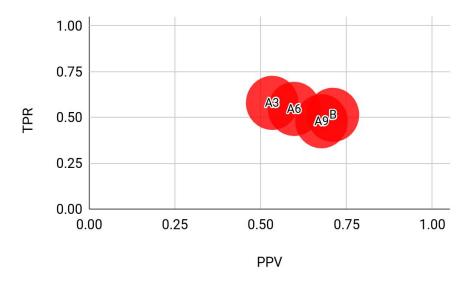
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- 4. Repeat for alphas.
- 5. Run b-matching procedure as before.

ALPHABETR Simulator

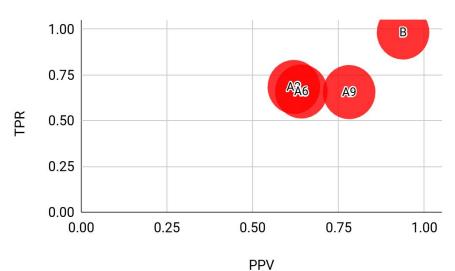
- Models PairSeq experiments
 - Number of clones, ~1000
 - Number of wells, 96 and 480
 - Dual alpha and dual beta rates, 0.3 and 0.06
 - Proportion of chains shared
- Has a few shortcomings:
 - Does not model T cell maturation, e.g. dual beta vs. dual alpha reuse
 - Is not real data.

Results - 480 wells

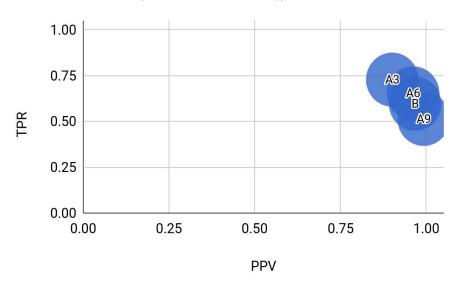
Clone identification (W=480, with sharing)



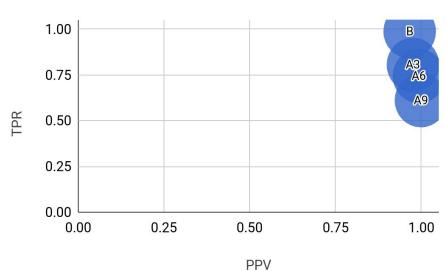
Clone identification (W=480, without sharing)



Pair identification (W=480, with sharing)

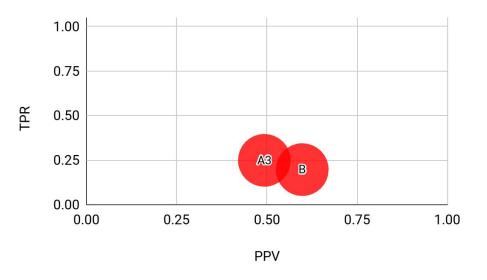


Pair identification (W=480, without sharing)

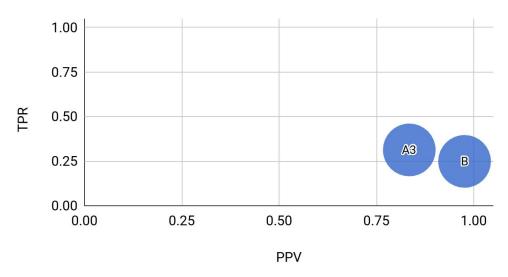


Results - 96 wells

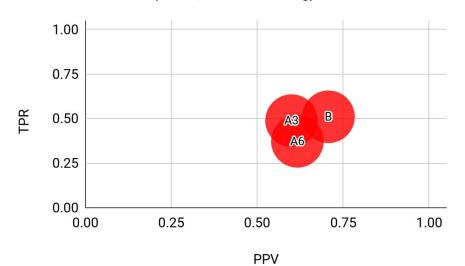
Clone identification (W=96, with sharing)



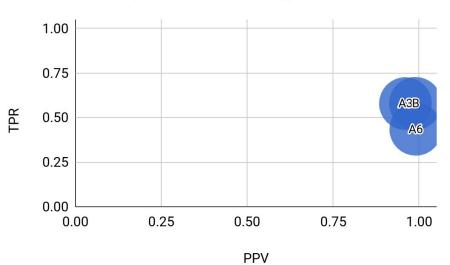
Pair identification (W=96, with sharing)



Clone identification (W=96, without sharing)

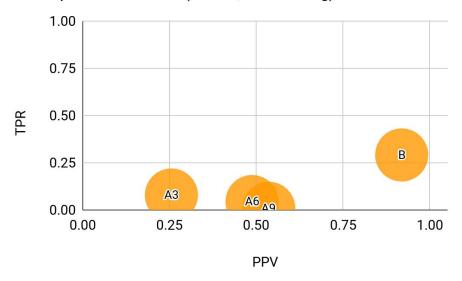


Pair identification (W=96, without sharing)

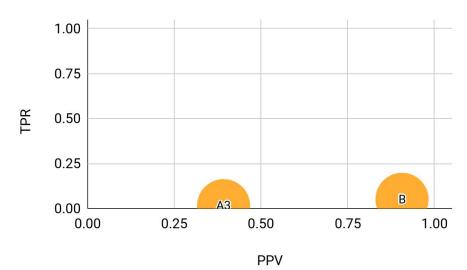


Results - Dual alphas

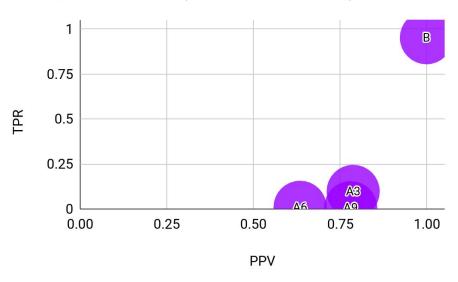
Dual alpha identification (W=480, with sharing)



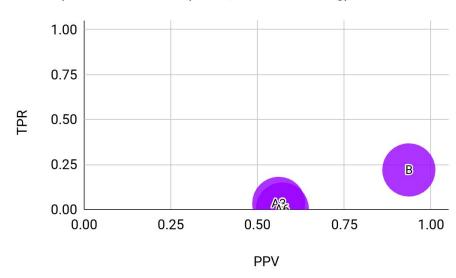
Dual alpha identification (W=96, with sharing)



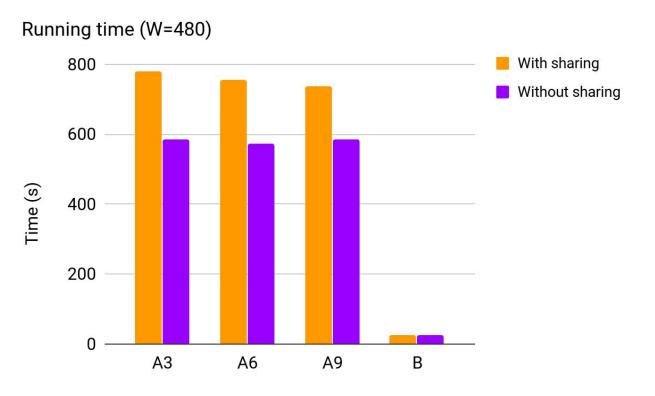
Dual alpha identification (W=480, without sharing)

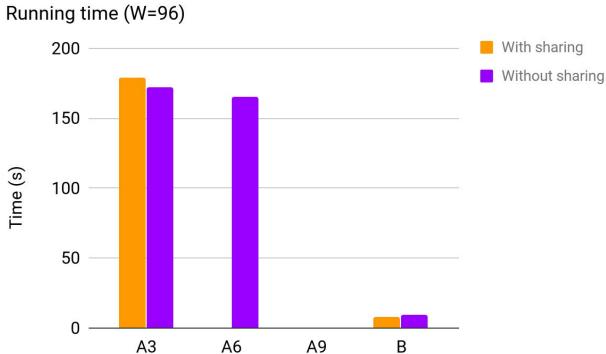


Dual alpha identification (W=96, without sharing)



Results - Running time





Results - b-matching dual identification

W = 480

PPV	With sharing	Without sharing
Dual alpha	0.9189	0.9985
Dual beta	0.7236	0.9870
Dual dual	0.8840	0.9624

TPR	With sharing	Without sharing
Dual alpha	0.2914	0.9523
Dual beta	0.4393	0.9446
Dual dual	0.1715	0.9493

W = 96

PPV	With sharing	Without sharing
Dual alpha	0.9065	0.9363
Dual beta	0.6462	0.6927
Dual dual	0.3800	0.7487

TPR	With sharing	Without sharing
Dual alpha	0.0508	0.2170
Dual beta	0.0676	0.2133
Dual dual	0.0219	0.1250

Future work

- 1. Incorporate chain-sharing into b-matching model.
- 2. Run b-matching procedure on real PairSeq data.
- 3. Generate and analyze larger datasets using ALPHABETR simulator.
- 4. Implement original PairSeq binomial model to analyze datasets.

Questions?